# PROCEEDINGS OF THE YEREVAN STATE UNIVERSITY

Chemistry and Biology

2013, № 2, p. 40–43

Biology

## OPTIMAL CONDITIONS OF INDUCTION OF L-AMINO ACID OXIDASE OF *ASPERGILLUS NIGER* R3 FUNGUS BY HYDROGEN PEROXIDE

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Our previous research has shown that L-amino acid oxidase activity in *Asp. niger* R3 yeast extracts was not detected and only some peroxisomal fractions have L-amino acid oxidase activity. Induction of L-amino acid oxidase is seen when hydrogen peroxide is added to the growth media. Our task was to investigate the induction of the enzyme by  $H_2O_2$  addition to incubation media. The results revealed that adding  $H_2O_2$  to *Asp. niger* R3 yeast extracts induce L-amino acid oxidase, especially it is more effective to use  $H_2O_2$  in the concentration of 0.001  $\mu M$  in the case of L-alanine as a substrate, and in the case of L-methionine – 0.003  $\mu M$  respectively.

*Keywords*: enzyme induction, L-alanine, L-methionine, deamination, inhibitory effect.

Introduction. According to the results obtained from our previous study about elaboration of induction of L-amino acid oxidase, addition of hydrogen peroxide in concentration 0.02 M to nutrient medium induces L-amino acid oxidase [1]. By increasing or decreasing the  $H_2O_2$  concentration, enzyme induction is expressed more poorly. It has been previously shown, that high concentrations of  $H_2O_2$  has inhibitory effect on the growth of S. cerevisiae yeasts, on the contrary low concentrations stimulate the growth [2]. It is interesting that static magnetic field and high power vibrations during first 20 min inhibit the growth of yeasts, but after 2 h stimulate the growth. According to the authors [3], static magnetic field and high power vibrations increase the level of hydrogen peroxide. It was shown, that extremely high magnetic field's rates (4 Hz) and hydrogen peroxide inhibit contractility of the heart by inducing Na-K ATP-ase, in the case when ultrasound stimulate ATP-ase activity. Authors concluded that  $H_2O_2$  is a messenger by which accomplished the electromagnetic field's action on the heart, and suppression of the heart contractility by ultrasound is due to the decrease in  $CO_2$  solubility [4]. Our previous investigations show, that there are low activity of L-amino acid oxidase in some peroxisomal fractions of Aspergillus Niger R3 [5]. According to

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the researchers [6] high concentrations of  $H_2O_2$  has cytotoxic effect, in the case when low concentrations induce many cellular functions, including enzyme induction. In order to make more effective induction of enzyme by  $H_2O_2$ , we investigated *Aspergillus Niger* R3 fungus's L-amino acid oxidase induction by adding  $H_2O_2$  not only to nutrient medium but also by adding to incubation medium for enzyme activity determination.

**Materials and Methods.** Aspergillus Niger R3 is served as research object, which is used in citric acid production. Rolen's synthetic nutrient media is used with composition of glucose – 2.0 g, KH<sub>2</sub>PO<sub>4</sub> – 0.05 g, MgSO<sub>4</sub>·7H<sub>2</sub>O – 0.05 g, ZnSO<sub>4</sub>·7H<sub>2</sub>O – 0.005 g, solved in 100 *Mm* tap water. For media growth we added 0.5 *ml* 33% H<sub>2</sub>O<sub>2</sub>. 0.4 g L-alanine was used as nitrogen source. Culture was made on 100 *ml* media (pH 7.0 42<sup>o</sup>C). 4 day grown culture *Aspergillus Niger* R3 at 32<sup>o</sup>C was homogenized in distilled water for 6 *min* in Potter-Elvedgiem's homogenizator [7]. To determine L-amino acid oxidase activity, enzyme preparation was incubated for 60 or 90 *min* in pyrophosphate buffer media in presence of corresponding substrates, L-alanine, L-methionine, L-valine and L-glutamate respectively, 10 *mM*. The reaction was stopped by 0.2% dinitrophenylhydrazine, then 0.1 *N* NaOH. Color intensity was measured at 440 *nm*. Enzyme activity was expressed in  $\mu$ M of produced ketoacid per 1 g of mycelium [8].

**Results and Discussion.** As it has been shown in our previous research (Tab. 1), in the absence of  $H_2O_2$  in the media the activity of L-amino acid oxidase was not detected, in the case of adding 0.02 *M* concentration of  $H_2O_2$  to the media the activity of L-amino acid oxidase was induced. Nevertheless, further increase of  $H_2O_2$  concentration decreased the activity of enzyme by 40%.

#### Table 1

Composition of nutrient media	L-amino acid oxidase activity (substrate L-alanine)	
without H <sub>2</sub> O <sub>2</sub>	0	
0.01 M H <sub>2</sub> O <sub>2</sub>	6.04±0.3	
$0.02 M H_2O_2$	9.5±0.4	
0.03 M H <sub>2</sub> O <sub>2</sub>	7.6±0.2	
0.045 <i>M</i> H <sub>2</sub> O <sub>2</sub>	6.1±0.2	

Effects of different  $H_2O_2$  concentrations in the growth media on L-amino acid oxidase activity in the extracts of Asp. niger R3 fungus ( $\mu$ M ketoacid per 1 g mycelium n = 5, p < 0.05)

It can be concluded, that  $H_2O_2$  over or less than 0.02 *M* concentration has inhibitory effect on enzyme activity. According to the results obtained from our previous investigations adding  $H_2O_2$  to the *Asp. niger* R3 yeast's growth media leads to the induction of L-amino acid oxidase.

In the next stage of our experiments we attempted to investigate the activity of L-amino acid oxidase by adding the  $H_2O_2$  to the incubation media. For that reason every 20 *min* 0.001 *M* concentration of  $H_2O_2$  was added to the incubation media. The results are presented in Tab. 2.

As one can notice from the results the activity of enzyme is not detected in the sample without  $H_2O_2$  in the incubation media. After 20 *min* incubation in the presence of 0.001  $\mu M$   $H_2O_2$  and L-alanine used as a substrate, the activity of

L-amino acid oxidase can be detected (4.45  $\mu M$  ketoacid/1 g mycelium). Further incubation for another 20 min with 0.002  $\mu M$  H<sub>2</sub>O<sub>2</sub> leads to a slight decrease in enzyme activity (1.9  $\mu M$  ketoacid/1 g mycelium). Another 20 min incubation with 0.003  $\mu M$  H<sub>2</sub>O<sub>2</sub> causes a dramatic decrease in enzyme activity (from 1.9  $\mu M$  to 0.6  $\mu M$  ketoacid / 1 g mycelium). The results obtained from the same experiment which has been done by using L-methionine instead of L-alanine as deamination substrate have some differences. It is notable, that the highest deamination activity in the case of using L-alanine as a substrate is noticed after first 20 min incubation by H<sub>2</sub>O<sub>2</sub> and the activity of enzyme when using L-methionine as a substrate equals 0 not only after first 20 min but also after 40 min incubation period. But, after 60 min incubation with 0.003  $\mu M$  H<sub>2</sub>O<sub>2</sub> L-amino acid oxidase activity increases to 2.8  $\mu M$  ketoacid / 1 g mycelium.

#### Table 2

Effect of different concentrations of  $H_2O_2$  in the incubation media on L-amino acid oxidase activity in the extracts of Asp. niger R3 yeasts (n = 5, p < 0.05)

Substrate	$H_2O_2$ concentrations,	Incubation	Enzyme activity
	$\mu M$	duration, min	$(\mu M \text{ ketoacid} / 1 g \text{ mycelium})$
L-alanine	without H <sub>2</sub> O <sub>2</sub>	-	0
	0.001	20	4.45 ±0.046
	0.002	40	$1.9 \pm 0.02$
	0.003	60	$0.6 \pm 0.01$
L-methionine	without H <sub>2</sub> O <sub>2</sub>	—	0
	0.001	20	0
	0.002	40	0
	0.003	60	$2.8 \pm 0.02$

It can be concluded, that  $H_2O_2$  stimulates L-amino acid oxidase activity by adding it not only to the growth media but also to the incubation media. It is interesting, that there are some differences in the pattern of L-amino acid oxidase activity induction by  $H_2O_2$  for different amino acids (L-alanine, L-methionine), and additional research is required for clarification.

Received 06.06.2013

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